

1 **Treatment with both nintedanib and steroid in bleomycin-induced pulmonary**

2 **fibrosis model mice exacerbated lung fibrosis**

3

4

Sadaaki Shiromori^a, Hirokuni Hirata^a, Kozo Sato^a, Kazuhiro Tada^a, Masafumi Arima^b,

5

Yasutsugu Fukushima^a

6

7

^aDepartment of Respiratory Medicine and Clinical Immunology, Dokkyo Medical

8

University Saitama Medical Center, 2-1-50 Minamikoshigaya, Koshigaya, Saitama

9

343-0845, Japan

10

^b Department of Rheumatology, Dokkyo Medical University, Tochigi, Japan.

11 Short Title: Nintedanib+steroid treatment exacerbates lung fibrosis

12

13

Corresponding Author:

14

Hirokuni Hirata, MD

15

Department of Respiratory Medicine and Clinical Immunology, Dokkyo Medical

16

University Saitama Medical Center, 2-1-50 Minamikoshigaya, Koshigaya, Saitama

17

343-0845, Japan

18

Tel: +81-48-965-1111, Fax: +81-48-965-1238, E-mail: hirokuni@dokkyomed.ac.jp

19

20 Keywords: nintedanib, steroid, bleomycin-induced pulmonary fibrosis

21

22

Abstract

23 **Background:** In the clinical setting, the optimal timing of administration of nintedanib
24 in the active or inactive phase of idiopathic pulmonary fibrosis (IPF) has not been
25 clarified. Also, no consideration has been made of synergistic actions with steroids.

26 **Objectives:** We used bleomycin-induced pulmonary fibrosis model mice to analyze the
27 effects of the timing of nintedanib administration and synergistic actions in combination
28 with steroid.

29 **Method:** In the bleomycin-induced pulmonary fibrosis mouse model, nintedanib was
30 given before or after bleomycin administration and the effects on pulmonary fibrosis
31 and fibroblast growth factors in bronchoalveolar lavage (BAL) fluid were analyzed.
32 Then, treatment with both steroid and nintedanib was administered in the same model
33 and the effects of combined use on lung fibrosis were analyzed. Severity of lung fibrosis
34 was analyzed using the Ashcroft score. Fibroblast growth factors in BAL fluid were
35 measured using Magnetic Luminex[®] assay and enzyme-linked immunosorbent assay.

36 **Results:** Pretreatment with nintedanib before administration of bleomycin, but not after
37 administration of bleomycin, reduced the Ashcroft score (2.4 ± 1.4 vs 4.9 ± 1.3 , $P < 0.01$)
38 and decreased TGF- β 1 in BAL fluid (95.7 ± 48.4 pg/mL vs 147.1 ± 11.7 pg/mL, $P < 0.05$).

39 Conversely, pretreatment with both steroid and nintedanib did not decrease the Ashcroft
40 score compared with pretreatment of saline as a control (Score: 3.8 ± 1.3 vs 4.6 ± 1.1).
41 There was also no synergistic effect of both steroid and nintedanib on pulmonary
42 fibrosis.

43 **Conclusions:** Nintedanib may be useful when administered early in patients with IPF.
44 Further, clinically, administration of combined nintedanib and steroid requires caution
45 in view of the effects on pulmonary fibrosis and side effects.

46

47

Introduction

48 Idiopathic pulmonary fibrosis (IPF) is one of the idiopathic interstitial pneumonias
49 (IIPs). It is a type of lung disease characterized by extensive fibrosis in the lungs and
50 causes restrictive ventilatory impairment. Compared with other IIPs, IPF shows poor
51 response to steroids and immunosuppressive drugs and carries a poor prognosis¹). IPF is
52 the most common of the IIPs.

53 The precise etiology is not yet known, but smoking is considered to be a risk factor
54 ²). In IPF, inflammation does not necessarily precede fibrosis, but with increased
55 damage to the alveolar epithelium by various stimuli, excessive collagen deposition

56 required for repair ensues with an abnormal repair reaction, leading to the progression
57 of fibrosis^{3,4}). Interstitial thickening is considered to reduce oxygen uptake causing
58 restrictive disorder (reduced vital capacity) due to decreased lung compliance;
59 symptoms include dry cough and dyspnea on exertion^{5,6}). Transforming growth factor
60 (TGF)- β is considered to play an important role in fibrosis, and it is known to cause
61 epithelial-mesenchymal transition (EMT) to type II alveolar epithelium^{7,8}). It is also
62 known to induce differentiation into fibroblasts and myofibroblasts⁹). There is no
63 curative treatment for IPF and currently available treatments, except for lung
64 transplantation, only slow progression¹⁰).

65 Recently, pirfenidone was reported to suppress fibrosis, and it has been found to
66 reduce deterioration in relation to forced vital capacity (FVC) and the 6-min walk test
67 compared with placebo^{11,12}). Although the mechanism of action of pirfenidone is
68 unclear, it is known to suppress TGF- β and tumor necrosis factor (TNF)- α in vitro¹³).

69 More recently, nintedanib, a small molecule tyrosine kinase inhibitor (TKI) and
70 indolinone derivative, has been used as a therapeutic agent for IPF^{14,15}). Vascular
71 endothelial growth factor (VEGF), fibroblast growth factor (FGF), and platelet-derived

72 growth factor (PDGF) promote angiogenesis and fibroblast proliferation ¹⁶⁻¹⁸.
73 Nintedanib acts on VEGF, FGF, and PDGF receptors ¹⁹) and thus inhibits PDGF, FGF,
74 VEGF-stimulated proliferation and migration of lung fibroblasts and subsequently
75 TGF- β -induced fibroblast transformation ^{19, 20}). However, in the clinical setting, the
76 timing of administration of nintedanib in the active or inactive phase of IPF has not
77 been clarified. Also, no consideration has been made regarding synergistic actions with
78 steroids.

79 Here, we used a bleomycin-induced pulmonary fibrosis mouse model to analyze
80 the effects of the timing of nintedanib administration and the synergistic action with
81 steroid.

82

83 **Materials and Methods**

84 *Animals*

85 Nine-week old female C57BL/6 mice were purchased from Japan SLC, Inc. (Shizuoka,
86 Japan). This study was approved by the Animal Ethics Committee of Dokkyo Medical
87 University Saitama Medical Center (No. 767).

88

89 ***Effects of treatment with nintedanib before and after administration of bleomycin***

90 To investigate the effect of nintedanib before and after inducing lung inflammation, we
91 performed transesophageal treatment with nintedanib before or after intratracheal
92 administration of bleomycin. Figure 1A shows the protocol. Saline (bleomycin[-])
93 (Group 1) or bleomycin sulfate (bleomycin[+]; 5 mg/kg body weight; Nippon Kayaku,
94 Tokyo, Japan) (Group 2, 3, and 4) was given by intratracheal administration to mice
95 anesthetized with medetomidine hydrochloride (Zenoaq, Fukushima, Japan), midazolam
96 (Astellas Pharma Inc., Tokyo, Japan), and butorphanol tartrate (Meiji Seika Pharma Co.,
97 Ltd., Tokyo, Japan), as a mixture of the 3 anesthetics (10 mL/kg body weight) on day
98 (D)1. Nintedanib (LC Laboratories, Boston, MA) (3 mg/kg body weight) with doses
99 similar to humans was given by transesophageal administration before (D0) (Group 3)
100 or after (D2) (Group 4) intratracheal administration of bleomycin. In contrast to
101 treatment with nintedanib, transesophageal saline was administered after (D2) (Group 2)
102 intratracheal administration of bleomycin. Subsequently, nintedanib (Group 1, Group 3,

103 and Group 4) or saline (Group 2) was given daily, twice a day from D2 to D6 and from
104 D8 to D13. The dose of nintedanib was determined similar to the dose used in humans.

105

106 ***Effects of treatment with both nintedanib and steroid before and after administration***
107 ***of bleomycin***

108 To investigate the effect of nintedanib with steroid before and after inducing lung
109 inflammation, we performed transesophageal treatment with nintedanib and
110 dexamethasone before or after intratracheal administration of bleomycin. Figure 2
111 shows the protocol. Groups 2, 3, and 4 were administered bleomycin to induce
112 pulmonary fibrosis. Saline (bleomycin[-]) (Group 1) or bleomycin sulfate
113 (bleomycin[+]; 5 mg/kg body weight) (Groups 2, 3, and 4) was given by intratracheal
114 administration to mice using a mixture of the 3 anesthetics on D1. Both nintedanib (3
115 mg/kg body weight) and moderate dexamethasone (0.1 mg/kg body weight) when
116 treated to humans was given by transesophageal administration before (D0) (Group 3)
117 or after (D2) (Group 4) intratracheal administration of bleomycin. Alternatively, saline
118 was given by transesophageal administration after (D2) (Group 2) intratracheal

119 administration of bleomycin. Subsequently, both nintedanib and dexamethasone
120 (Groups 1, 3, and 4) or saline (Group 2) was given daily, twice and once a day,
121 respectively, from D2 to D6 and from D8 to D13.

122

123 ***Bronchoalveolar lavage***

124 A single incision was made in the neck and the tissue covering the trachea was snipped
125 to expose the tracheal rings. Bronchoalveolar lavage (BAL) fluid was collected on D15
126 after bleomycin or saline administration; histological examination of the lungs was also
127 performed on D15. Using a 1.5 ml syringe, 0.5 ml was injected into the trachea once,
128 and BAL was performed 3 times. The BAL fluid sample was centrifuged at 400 g for
129 10 min at 4°C, and the supernatant was stored at -80°C until analysis.

130

131 ***Concentration of FGF2, PDGF-AA, VEGF-A, and collagen in BAL fluid***

132 BAL fluid samples were concentrated by centrifugal ultrafiltration (Amicon Darmstadt,
133 Germany), which is used to concentrate low molecular weight components. The cut-off
134 value for molecular weight was 3000. BAL fluid levels of FGF2, PDGF-AA, VEGF-A

135 were measured by Magnetic Luminex Assay, Mouse Premixed Multi-Analyte Kit (R&D,
136 Minneapolis, MN) using a multi-item simultaneous measurement device (Luminex
137 Japan Corporation, Tokyo, Japan). The minimum detectable dose (MDD) was 41.3
138 pg/mL, 0.9 pg/mL, and 4.0 pg/mL, respectively. In addition, concentrations of TGF- β 1
139 were measured using enzyme-linked immunosorbent assay kits (R&D, Minneapolis,
140 MN) according to the manufacturer's instructions. In preliminary experiment, albumin
141 concentration in BAL was measured for 5 mice in all groups presented this time, and
142 corrected for the concentration of each protein (TGF- β , VEGF, PDGF-AA). However,
143 since there was no significant difference, no correction by albumin concentration was
144 performed in this study. The MDD was 4.6 pg/mL. Collagen measurements were
145 determined using the Sircol Collagen Assay kit (Biocolor Ltd., Belfast, UK)²¹. The
146 collagen concentration was measured at 549 nm on a spectrophotometer.

147

148 ***Histopathological analysis***

149 The right lung tissues were preserved in order to develop this study in the future, and
150 the left lung was used in this study. To fix the lung, paraformaldehyde in phosphate

151 buffer was perfused into the left lung at a constant pressure. After fixation, lung tissues
152 were embedded in paraffin and sectioned with a microtome at a thickness of 3-5 μm .
153 Sections were mounted on slides and stained with hematoxylin and eosin (HE),
154 Elastica-Masson (EM), and for immunohistochemistry (IHC). For each mouse, three
155 sections of the whole lung stained with HE and EM were selected randomly. To analyze
156 the severity of lung fibrosis, grading was scored on a scale from 0 to 8, using the
157 average of microscope field scores, as described previously ²²). Biological micrographs
158 were taken with a digital camera for microscope (model DP73; Olympus Corporation,
159 Tokyo, Japan) with a $\times 10$ and $\times 40$ lens.

160

161 ***Immunostaining analysis***

162 Paraffin-embedded sections of tissue were deparaffinized and dehydrated following
163 antigen retrieval with citric acid buffer, as described previously ²³). Slides were first
164 incubated with blocking IgG solution for 1 h and then overnight with rabbit
165 anti-TGF- $\beta 1$ antibody (1:100 dilution; Abcam), rabbit anti-FGF2 antibody (1:200
166 dilution; Bioss, Woburn, MA), anti-VEGF-A antibody (1:200 dilution; Abcam), or

167 rabbit anti-PDGF-AA antibody (1:400 dilution; Bioss). The secondary antibody used
168 was a labeled polymer containing peroxidase and Fab' anti-IgG bound to an amino acid
169 polymer (Histofine mouse stain kit; Nichirei Bioscience, Tokyo, Japan).

170

171 ***Statistical analysis***

172 Data are expressed as means \pm standard deviation (SD). Statistical significance was
173 determined with the Mann-Whitney *U* test for each group. *P* values < 0.05 were
174 considered significant. JMP software (Version 11.0; SAS Institute, Cary, NY) was used
175 for statistical analyses.

176

177 ***Results***

178 ***Effects of nintedanib before and after administration of bleomycin: histopathological***
179 ***analysis and TGF- β 1, FGF2, PDGF-AA, VEGF-A, and collagen concentrations in***
180 ***BAL fluid***

181 We first performed histopathological analysis (Figure. 3A to Figure. 3H) and measured
182 concentrations of collagen (Figure. 3J), TGF- β 1 (Figure. 3K), PDGF-AA (Figure. 3L),

183 and VEGF-A (Figure. 3M) in BAL fluid on D15 of lung treatment with nintedanib
184 before and after administration of bleomycin in. The Ashcroft score as an index of lung
185 fibrosis (Figure. 3I) was significantly reduced in Group 3 compared with Group 2
186 (2.4 ± 1.4 vs 4.9 ± 1.3 , $P < 0.01$) but not in Group 4 (3.6 ± 1.5 , $P = 0.106$) compared with
187 Group 2.

188 The concentration of collagen in BAL fluid was significantly reduced in Group 3
189 compared with Group 2 (4.4 ± 3.0 $\mu\text{g/mL}$ vs 10.0 ± 6.5 $\mu\text{g/mL}$, $P < 0.05$; Figure. 3J) but not
190 in Group 4 (7.0 ± 3.0 pg/mL) compared with Group 2. In addition, the concentration of
191 TGF- β 1 in BAL fluid was significantly reduced in Group 3 compared with Group 2
192 (95.7 ± 48.4 pg/mL vs 147.1 ± 11.7 pg/mL , $P < 0.05$; Figure. 3K) but not in Group 4
193 (120.2 ± 26.4 pg/mL) compared with Group 2. The concentrations of PDGF-AA and
194 VEGF-A in BAL fluid were not reduced in either Group 4 or Group 3 compared with
195 Group 2. FGF2 was not detected in BAL fluid in any of the groups (data not shown).

196

197 ***Bleomycin-induced pulmonary fibrosis: IHC analysis***

198 We performed IHC analysis on D15 after administration of bleomycin (Figure 4).
199 TGF- β 1 were expressed in lung fibroblasts (Figure 4B). Although FGF2 was not
200 detected in BAL fluid, it was expressed in lung fibroblasts (Figure 4D). PDGF-AA
201 (Figure 4F) and VEGF-A (Figure 4H) were expressed mainly in endothelial and
202 epithelial cells.

203

204 *Effects of both steroid and nintedanib before and after administration of bleomycin:*

205 *Histopathological analysis, and collagen concentrations in BAL fluid*

206

207 Finally, we performed histopathological analysis on D15 of treatment with both steroid
208 and nintedanib before and after administration of bleomycin (Figure. 5A to Figure.5H).
209 Ashcroft score (Figure. 5I) was not reduced in Group 3 (3.8 ± 1.3) or Group 4 (4.1 ± 1.3)
210 compared with Group 2 (4.6 ± 1.1). In addition, concentration of collagen (Figure. 5J) in
211 BAL fluid was not reduced in Group 3 (11.1 ± 4.2) or Group 4 (10.2 ± 4.0) compared with
212 Group 2 (12.3 ± 6.8).

213

214 **Discussion**

215 In this study, pretreatment of mice with pulmonary fibrosis with nintedanib before
216 administration of bleomycin, but not after administration of bleomycin, reduced the
217 Ashcroft score, decreased collagen and TGF- β 1 in BAL fluid was seen to be associated
218 with a decrease in lung fibrosis. The pretreatment with both nintedanib and steroid
219 exacerbated lung fibrosis in bleomycin-induced pulmonary fibrosis model mice
220 compared with pretreatment with nintedanib only.

221 Bleomycin-induced lung fibrosis is a widely used animal model of pulmonary
222 fibrosis ²⁴). Intratracheal administration of bleomycin induces acute alveolitis and
223 interstitial inflammation, characterized by sequential recruitment of leucocytes in the
224 first week, followed by fibrotic responses associated with fibroblast proliferation and
225 synthesis of extracellular matrix in the second week ²⁵). Steroids have
226 anti-inflammatory effects and nintedanib has anti-fibrotic effects ^{19, 26}). We
227 demonstrated the effect of nintedanib, which has anti-fibrotic function, in mice with
228 bleomycin-induced pulmonary fibrosis and then showed that bleomycin-induced
229 pulmonary fibrosis was reduced by pretreatment with nintedanib before administration

230 of bleomycin, but not by treatment with nintedanib after administration of bleomycin.

231 The results were similar to those reported by Lutz et al. ²⁷⁾. Fibroblasts are thought to

232 proliferate once activated and this cannot be suppressed. However, we demonstrated

233 that the concentration of TGF- β 1 in BAL fluid decreased with the extent of lung fibrosis.

234 Conversely, FGF2 was not detected despite the presence of lung fibrosis. However, IHC

235 of lung tissues showed clearly that FGF2 was also expressed in fibroblasts. Depending

236 on the type, cytokines may not be detected in BAL. In addition, nintedanib had no

237 effect on PDGF-AA and VEGF-A in BAL fluid. PDGF and VEGF can be produced by

238 fibroblasts, but they are also produced in large amounts by endothelial cells, epithelial

239 cells, and macrophages ^{28, 29)}. This suggests that even if both the activity and

240 proliferation of fibroblasts was suppressed by nintedanib, the cytokine concentration in

241 BAL was not affected. In addition, the treatment of nintedanib before and after

242 administration with bleomycin had no effect on the number of total cells, including

243 macrophages, lymphocytes, neutrophils and eosinophils in BAL (data not shown).

244 Clinically, it is desirable to begin administering nintedanib from the period when

245 fibroblast activity is as low as possible. Nintedanib, which is a TKI that acts via VEGF,

246 PDGF, and FGF receptors expressed on lung fibroblasts, may have suppressed
247 subsequent TGF- β 1 and FGF2 production by directly suppressing fibroblast activity and
248 proliferation, regardless of inflammatory cells. In addition, the single treatment with
249 nintedanib before administration of bleomycin without the subsequent treatment with
250 nintedanib after administration of bleomycin did not suppress pulmonary fibrosis (data
251 not shown). This result indicates that treatment with nintedanib is optimal by continuous
252 administration.

253 When treating pulmonary fibrosis, it is important to control inflammation and
254 subsequent fibrosis. Steroids act on lymphocytes and macrophages to suppress
255 inflammatory cytokines and exert anti-inflammatory effects ^{30, 31}). However, there are
256 many negative reports about the effectiveness of steroids in ameliorating human IPF ^{32,}
257 ³³). In addition, studies in murine models showed that treatment with steroids did not
258 suppress lung fibrosis ^{34, 35}). The effectiveness of combining steroids with antifibrotic
259 drugs in human IPF is also unclear. For this reason, we analyzed the effectiveness of a
260 human antifibrotic drug and steroid in similar amounts using the bleomycin mouse
261 model. However, the results showed that the combinations of various nintedanib doses

262 and steroid could not suppress pulmonary fibrosis. Thus, pulmonary fibrosis aggravated
263 by the single-agent administration of nintedanib was observed to be aggravated by the
264 combined use of both agents after the administration of bleomycin, regardless of
265 pretreatment with steroid. These findings indicate that this combined use after the
266 induction of inflammation may not inhibit pulmonary fibrosis. Although the detailed
267 mechanism is unknown, it is considered that the steroid may reduce the affinity of
268 nintedanib for fibroblasts or this may be reduced by interaction between the drugs. In
269 addition, although the dose of steroid was relatively high in this study, the effect on
270 pulmonary fibrosis may be affected by the administration of a small dose of steroid.

271 In conclusion, nintedanib may be useful when administered early in patients with
272 pulmonary fibrosis. Further, clinically, it is considered that a combination of nintedanib
273 and steroid should be used with sufficient caution from the viewpoint of effects on
274 pulmonary fibrosis and side effects.

275

276

Acknowledgements

277

278

279

The authors would like to thank S. Sekiguchi, S. Ohno, K. Fukuda, K. Tsujikado, A. Yamamoto, Y. Eda, A. Hara, H. Tanaka, and M. Nomaguchi at the Department of Respiratory Medicine and Clinical Immunology, Dokkyo Medical University Saitama

280 Medical Center, for their excellent technical assistance.

281

282

Conflict of interest:

283

All authors have no conflict of interest.

284

285

Abbreviations:

286 IPF: idiopathic pulmonary fibrosis, IIPs: idiopathic interstitial pneumonias, TGF:

287 transforming growth factor, EMT: epithelial-mesenchymal transition, FVC: forced vital

288 capacity, TNF: tumor necrosis factor, TKI: tyrosine kinase inhibitors, VEGF: Vascular

289 endothelial growth factor, PDGF: platelet-derived growth factor, FGF: fibroblast growth

290 factor, MDD: minimum detectable dose, BAL: bronchoalveolar lavage, HE:

291 hematoxylin and eosin, EM: Elastica-Masson

292

293 **References**

294 1. Scriabine A, Rabin DU: New developments in the therapy of pulmonary fibrosis

295 Adv Pharmacol **57** : 419-464, 2009.

296 2. Ballester B, Milara J, Cortijo J: Idiopathic pulmonary fibrosis and lung cancer:

297 mechanisms and molecular targets. Int J Mol Sci **20** : 593, 2019.

- 298 3. Ohkouchi S, Ono M, Kobayashi M, et al: Myriad functions of stanniocalcin-1
299 (STC1) cover multiple therapeutic targets in the complicated pathogenesis of
300 idiopathic pulmonary fibrosis (IPF). *Clin Med Insights Circ Respir Pulm Med*
301 **9**(Suppl 1) : 91-96, 2015.
- 302 4. Selman M, Thannickal VJ, Pardo A, et al: Idiopathic pulmonary fibrosis:
303 pathogenesis and therapeutic approaches. *Drugs* **64** : 405-430, 2004.
- 304 5. Vigeland CL, Hughes AH, Horton MR: Etiology and treatment of cough in
305 idiopathic pulmonary fibrosis. *Respir Med* **123** : 98-104, 2017.
- 306 6. Miki K, Maekura R, Miki M, et al: Exertional acidotic responses in idiopathic
307 pulmonary fibrosis: The mechanisms of exertional dyspnea. *Respir Physiol*
308 *Neurobiol* **185** : 653-658, 2013.
- 309 7. Felton VM, Borok Z, Willis BC: N-acetylcysteine inhibits alveolar
310 epithelial-mesenchymal transition. *Am J Physiol Lung Cell Mol Physiol* **297** :
311 L805-12, 2009.
- 312 8. Willis BC, Borok Z: TGF-beta-induced EMT: Mechanisms and implications for
313 fibrotic lung disease. *Am J Physiol Lung Cell Mol Physiol* **293** : L525-534, 2007.

- 314 9. Upagupta C, Shimbori C, Alsilmi R, et al: Matrix abnormalities in pulmonary
315 fibrosis. *Eur Respir Rev* **27** : 180033, 2018.
- 316 10. George PM, Patterson CM, Reed AK, et al: Lung transplantation for idiopathic
317 pulmonary fibrosis. *Lancet Respir Med* **7** : 271-282, 2019.
- 318 11. Xaubet A, Serrano-Mollar A, Ancochea J: Pirfenidone for the treatment of idiopathic
319 pulmonary fibrosis. *Expert Opin Pharmacother* **15** : 275-281, 2014.
- 320 12. Cottin V: Changing the idiopathic pulmonary fibrosis treatment approach and
321 improving patient outcomes. *Eur Respir Rev* **21** : 161-167. 2012.
- 322 13. Maher TM: Pirfenidone in idiopathic pulmonary fibrosis. *Drugs Today (Barc)* **46** :
323 473-482, 2010.
- 324 14. Varone F, Sgalla G, Iovene B, et al: Nintedanib for the treatment of idiopathic
325 pulmonary fibrosis. *Expert Opin Pharmacother* **19** : 167-175, 2018.
- 326 15. Wollin L, Distler JHW, Redente EF, et al: Potential of nintedanib in treatment of
327 progressive fibrosing interstitial lung diseases. *Eur Respir J* **54** : 1900161, 2019.

- 328 16. Naim R, Chang RC, Sadick H, et al. Effect of vascular endothelial growth factor on
329 fibroblasts from external auditory canal cholesteatoma. Arch Med Res **36** : 518-523,
330 2005.
- 331 17. Grazul-Bilska AT, Luthra G, Reynolds LP, et al: Effects of basic fibroblast growth
332 factor (FGF-2) on proliferation of human skin fibroblasts in type II diabetes mellitus.
333 Exp Clin Endocrinol Diabetes. **110** : 176-181, 2002.
- 334 18. Lu J, Shi J, Gui B, et al: Activation of PPAR- γ inhibits PDGF-induced proliferation
335 of mouse renal fibroblasts. Eur J Pharmacol **789** : 222-228, 2016.
- 336 19. Wind S, Schmid U, Freiwald M, et al: Clinical pharmacokinetics and
337 pharmacodynamics of nintedanib. Pharmacokinet **58** : 1131-1147, 2019.
- 338 20. Prashanth Goud M, Bale S, Pulivendala G, et al: Therapeutic effects of nimbolide,
339 an autophagy regulator, in ameliorating pulmonary fibrosis through attenuation of
340 TGF- β 1 driven epithelial-to-mesenchymal transition. Int Immunopharmacol **75** :
341 105755, 2019.
- 342 21. Tokuda A, Itakura M, Onai N, et al: Pivotal role of CCR1-positive leukocytes in
343 bleomycin-induced lung fibrosis in mice. J Immunol **164** : 2745-2751, 2000.

- 344 22. Ashcroft T, Simpson JM, Timbrell V: Simple method of estimating severity of
345 pulmonary fibrosis on a numerical scale. *J Clin Pathol* **41** : 467-70, 1998.
- 346 23. Henderson NC, Mackinnon AC, Farnworth SL, et al: Galectin-3 expression and
347 secretion links macrophages to the promotion of renal fibrosis. *Am J Pathol* **172** :
348 288-298, 2008.
- 349 24. Walters DM, Kleeberger SR: Mouse models of bleomycin-induced pulmonary
350 fibrosis. *Curr Protoc Pharmacol* **Mar;Chapter 5**:Unit 5.46, 2008.
- 351 25. Monica L, Silvia F, Enrico S, et al: Ajulemic acid exerts potent anti-fibrotic effect
352 during the fibrogenic phase of bleomycin lung. *Respir Res* **17** : 49, 2016.
- 353 26. Khan MO, Lee HJ: Synthesis and pharmacology of anti-Inflammatory steroidal
354 anti-drugs. *Chem Rev* **108** : 5131-5145, 2008.
- 355 27. Lutz W, Isabelle M, Valérie Q, et al: Antifibrotic and anti-inflammatory activity of
356 the tyrosine kinase inhibitor nintedanib in experimental models of lung fibrosis. *J*
357 *Pharmacol Exp Ther* **349** : 209-220, 2014.
- 358 28. Coultas L, Chawengsaksophak K, Rossant J: Endothelial cells and VEGF in
359 vascular development. *Nature* **438** : 937-945, 2005.

- 360 29. James CB. Regulation of PDGF and its receptors in fibrotic diseases. *Cytokine &*
361 *Growth Factor Reviews* **15** : 255-273, 2004.
- 362 30. Guo-W , Yi S, Yi-Jun Z, et al: Glucocorticoid attenuates acute lung injury through
363 induction of type 2 macrophage. *J Transl Med* **15** : 181, 2017.
- 364 31. Barnes PJ. Anti-inflammatory actions of glucocorticoids: Molecular mechanisms.
365 *Clin Sci (Lond)* **94** : 557-572, 1998.
- 366 32. Gay SE, Kazerooni EA, Toews GB, et al: Idiopathic pulmonary fibrosis: Predicting
367 response to therapy and survival. *Am J Respir Crit Care Med* **157** : 1063-1072,
368 1998.
- 369 33. Flaherty KR, Toews GB, Lynch JP, et al: Steroids in idiopathic pulmonary fibrosis:
370 A prospective assessment of adverse reactions, response to therapy, and survival.
371 *Am J Med* **110** : 278-282, 2001.
- 372 34. Hosoya T: Steroid resistance and lung-tissue cytokines in experimental
373 bleomycin-induced lung fibrosis. *Nihon Kyobu Shikkan Gakkai Zasshi* **35** : 766-775,
374 1997.

375 35. Tamagawa K, Taooka Y, Maeda A, et al: Inhibitory effects of a lecithinized
376 superoxide dismutase on bleomycin-induced pulmonary fibrosis in mice. Am J
377 Respir Crit Care Med **161** : 1279-1284, 2000.

378

379 **Figure legends**

380 Figure 1; Effects of treatment with nintedanib before and after administration of
381 bleomycin in mice. Saline (bleomycin[-]) (Group 1) or bleomycin sulfate
382 (bleomycin[+]; 5 mg/kg body weight (Groups 2, 3, and 4) was given by intratracheal
383 administration to mice on D1. Nintedanib (3 mg/kg body weight) was given by
384 transesophageal administration before (D0) (Group 3) or after (D2) (Group 4)
385 intratracheal administration of bleomycin in mice. As a control, saline was given by
386 transesophageal administration after (D2) (Group 2) intratracheal administration of
387 bleomycin. Subsequently, nintedanib (Groups 1, 3, and 4) or saline (Group 2) was
388 administered daily, twice a day from D2 to D6 and from D8 to D13. Nint; nintedanib,
389 BLM; bleomycin, BALF; bronchoalveolar lavage fluid, ELISA; Enzyme-Linked
390 Immuno Sorbent Assay.

391

392 Figure 2; Effects of treatment with both nintedanib and steroid before and after
393 administration of bleomycin in mice. Figure 2 shows the study protocols. Group 2, 3,
394 and 4 were administered bleomycin to induce pulmonary fibrosis. Saline (bleomycin[-])
395 (Group 1) or bleomycin sulfate (bleomycin[+]; 5 mg/kg body weight) (Group 2, 3, and
396 4) was given by intratracheal administration to mice on D1. Both nintedanib (3 mg/kg
397 body weight) and dexamethasone (0.1 mg/kg body weight) was given by
398 transesophageal administration before (D0) (Group 3) or after (D2) (Group 4)
399 intratracheal administration of bleomycin. As a control, saline was given by
400 transesophageal administration after (D2) (Group 2) intratracheal administration of
401 bleomycin. Subsequently, both nintedanib and dexamethasone (Groups 1, 3, and 4) or
402 saline (Group 2) were given daily, twice and once a day, respectively, from D2 to D6
403 and from D8 to D13. Nint; nintedanib, Dex; dexamethasone, BLM; bleomycin, BALF;
404 bronchoalveolar lavage fluid.

405

406

407 Figure 3; Histopathological analysis [A to I] and collagen concentrations [J], TGF- β 1
408 [K], VEGF-A [L], and PDGF-AA [M] in BAL fluid on D15 to determine the effects of
409 nintedanib before and after administration of bleomycin in mice. For histopathological
410 analysis, lung sections were stained with HE [A, C, E, G] and EM [B, D, F, H]. Group1;
411 [A], [B], Group2; [C], [D], Group3; [E], [F], Group4; [G], [H]. Scale bar: 100 μ m.
412 Quantification of lung fibrosis [I] in lung tissue by score. Concentrations (ug/ml) of
413 collagen in BAL fluid [J]. P values indicate comparisons between each group. Data are
414 means \pm SD for each of 6 to 11 mice. NS, non-significance. N.D, not detected.

415

416 Figure 4; Immunostaining analysis (TGF- β 1 [A], FGF2 [C], PDGF-AA [E], and
417 VEGF-A [G]) on D15 after administration of bleomycin. Scale bar: 20 μ m, or 50 μ m. As
418 controls, immunostaining analysis of TGF- β 1 [B], FGF2 [D], PDGF-AA [F], and
419 VEGF [H] is shown as negative results in which primary antibody were absent.

420

421 Figure 5; Histopathological analysis [A to I] and collagen concentrations [J] in BAL
422 fluid on D15 for the effects of both steroid and nintedanib before and after

423 administration of bleomycin. For histopathological analysis, lung sections were stained
424 with HE [A, C, E, G] and EM [B, D, F, H]. Group1; [A], [B], Group2; [C], [D],
425 Group3; [E], [F], Group4; [G], [H]. Scale bar: 100 μ m. Quantification of lung fibrosis
426 [I] in lung tissue by score. Concentrations (ug/ml) of collagen in BAL fluid [J]. Data are
427 means \pm SD for each of the 9 to 11 mice. NS, non-significant.