

node with unknown primary origin. The patient developed multiple cranial neuropathies and sensory neuropathy, which presented as acute bulbar palsy.

Clinical Summary

A 65-year-old man developed discomfort when swallowing from the end of August 2020. After a few days, he was unable to drink water and consulted an otolaryngologist. There were no findings of passage obstruction, so a neurological disease was suspected, and the patient was referred to our hospital. The medical history of the patient was significant for hypertension, gastric ulcer, colon polyp, and cervical spondylosis. He smoked 20 cigarettes a day for 45 years. On admission, he was alert and had dysphagia, diminished pharyngeal muscle reflex, and disappearance of soft palate reflex. No obvious tongue atrophy or fasciculation was observed. Symmetric numbness was observed in the periphery of fingers. The triceps tendon reflex was weak, whereas the others were absent in both upper and lower limbs. Blood thiamine (37 ng/mL; normal range: 24-66), mecobalamin (237 pg/mL; normal range: 180-914), and folic acid (6.4 ng/mL; lower limit: 4.0) levels were in the normal range. ANCA-related autoantibodies, anti-AChR antibody, anti-MuSK antibody, anti-GM1 IgM and IgG antibodies, and anti-GQ1b IgM and IgG antibodies were negative. There was no decrease in the cerebrospinal fluid (CSF)/serum glucose ratio³ and the IgG index was 0.819 (i.e., slightly elevated)⁴. CSF showed class I cytopathology (Table 1). The head and cervical spine MRI showed no abnormalities. The nerve conduction test showed no decrease in conduction velocity or conduction block. Based on the acute clinical course and laboratory findings, acute bulbar palsy (ABP)^{5,6}, a subtype of Guillain-Barré syndrome, was suspected and two courses of IVIg were administered. However, the symptoms did not improve. Anti-GT1a antibody, which is a marker of ABP, was negative (Table 2). On the 36th day of hospitalization, right abducens, facial, and hypoglossal nerve palsies appeared, and on the 50th day of hospitalization, he required intubation and mechanical ventilation for respiratory failure. The patient was investigated to rule out malignant lymphoma and PNS as the causes of his symptoms. No abnormal findings were found on random skin biopsy or whole-body CT. We were unable to

conduct a PET study at our facility. The laboratory findings showed a slight increase in sIL2R, CEA, and SCC levels. The anti-Hu antibody was positive. Progastrin-releasing peptide (ProGRP) and neuron-specific enolase (NSE) specific to small cell carcinoma were not elevated initially but became significantly elevated at the later stage (ProGRP: 111 pg/ml). Although no pulmonary lesion was observed on chest CT, progressive enlargement of the lymph node near the para-aortic lymph node was observed at about 3 months (Fig. 1). Lymph node excision was performed at that site using video-assisted thoracic surgery, which led to the diagnosis of PNS associated with small cell carcinoma (Fig. 2). No chemotherapy or radiation therapy was administered after excision and 16 months after the resection, no new lesions were found on chest CT scan. In addition, the patient was alive without exacerbation of his neurological symptoms.

Discussion

PNS is a rare disease that affects less than 0.01% of cancer patients¹. More than half of the patients have neurological symptoms due to the tumor, and most tumors are diagnosed within 2 years of onset of neurological symptoms. The neural antibodies are detected in less than 50% of patients². The diagnosis is strongly suspected if the treatment of the tumor improves the neurological symptoms. The 2021 diagnostic criteria for PNS-associated antibodies are as follows: high-risk antibodies with more than 70% tumor complications, intermediate-risk antibodies with 30-70% tumor complications, and low-risk antibodies with less than 30% tumor complications (i.e., essential autoimmune diseases)⁷. High-risk antibodies typically present with subacute cerebellar ataxia, limbic encephalitis, encephalomyelitis, and subacute sensory ataxic neuropathy. In our patient, high-risk antibodies (i.e., anti-Hu antibody) presented with ABP, oculomotor palsy, and peripheral neuropathy.

ABP was initially suspected and treatment was initiated, but the patient did not improve. Subsequently, the patient developed new multiple cranial nerve palsies of late onset, suggesting that the disease course was different than that in ABP. The isolated lymph node near the aortic arch enlarged over time, which led to the diagnosis of the tumor. Hu antigen is found

Table 1 Laboratory Findings at the Admission.

<i>Hematology</i>		TP	7.2 g/dL	<i>Serology</i>	<i>Cerebrospinal fluid</i>
WBC	7.1 × 10 ⁹ /L	Albumin	4.28 g/dL	CRP	Cell
RBC	4.42 × 10 ¹² /L	UN	22 mg/dL	FT4	TP
Hb	14.3 g/dL	Na	143 mmol/L	FT3	Glucose
Ht	42.9 %	K	4.6 mmol/L	TSH	CSF/serum glucose ratio
MCV	97.1 fL	Cl	108 mmol/L	IgG	Albumin
MCH	32.4 pg	UA	13.6 mg/dL	CEA	IgG
MCHC	33.30 %	Creatinine	0.89 mg/dL	SCC	ADA
Platelet	23.8 × 10 ⁴ /μL	Glucose	114 mg/dL	ProGRP	MBP
<i>Coagulation</i>		CPK	60 U/L	NSE	sIL2R
PT-INR	0.98	HbA1c	5.0 %	sIL2 receptor	OB
APTT	30.3 sec	TG	102 mg/dL	PR3-ANCA	IgG index
D-dimer	0.63 μg/mL	HDL-C	45 mg/dL	MPO-ANCA	Cytology
<i>Biochemistry</i>		LDL-C	115 mg/dL	CLβ2GPI Ab	
AST	19 U/L	Thiamine	37 ng/mL	ACE	
ALT	15 U/L	Mecobalamin	237 pg/mL	ADA	
ALP	143 U/L	Folic acid	6.4 ng/mL	Anti-AchR Ab	
LDH	109 U/L			Anti-MuSK Ab	
GGT	21 U/L			Anti-Hu Ab	
T-Bil	1.20 mg/dL			3 +	

Abbreviations: ACE, angiotensin converting enzyme; Anti-AchR Ab, anti-acetylcholine receptor antibody; Anti-Hu Ab, antineuronal nuclear antibody; Anti-MuSK Ab, anti-muscle-specific tyrosine kinase; APTT, activated partial thromboplastin time; DA, adenosine deaminase; AST, aspartate aminotransferase, ALT: alanine aminotransferase; BUN, blood urea nitrogen; CEA, carcinoembryonic antigen; GGT, γ-glutamyltranspeptidase; Cl, chloride; CPK, creatine phosphokinase; CRP, C-reactive protein; Hb, hemoglobin; HDL-C, high density lipoprotein cholesterol; Ht, hematocrit; K, potassium; LDH, lactate dehydrogenase; LDL-C, low-density lipoprotein cholesterol; MBP, myeline basic protein; MCV, mean corpuscular volume; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; MPO-ANCA, myeloperoxidase-anti-neutrophil cytoplasmic antibodies; Na, sodium; NAG, N-acetyl-β-Dglucosaminidase; NSE, neuron-specific enolase; OB, Oligoclonal band IgG; Plt, platelet; PR3-ANCA, proteinase-3-anti-neutrophil cytoplasmic antibodies; ProGRP, pro-gastrin releasing peptide; PT-INR, prothrombin time-international normalized ratio; RBC, red blood cell; SCC, squamous cell carcinoma antigen; sIL2 receptor, soluble interleukin 2 receptor; T-Bil, Total Bilirubin; TG, triglyceride; TP, total protein; UA, uric acid. Bold were abnormal findings.

in the nucleus and cytoplasm of neurons in the cerebrum, spinal cord, and dorsal root ganglia; this antigen

is believed to be involved in cell development and differentiation.

Anti-Hu antibody positivity is associated with a high rate of SCLC⁸). Although neurological symptoms were observed before the diagnosis of tumor, no pulmonary lesions other than lymphadenopathy were observed during the disease. Several cases of anti-Hu antibody-positive PNS with lymph node metastasis of unknown cause have been reported in Japan^{9,10}. Neurologic findings in these patients included abnormal sensations in the upper extremities, diminished tendon reflexes, and normal nerve conduction studies. Our patient had sensory neuronopathy due to positive anti-Hu antibodies. The treatment of PNS begins with the treatment of the causative tumor and early treatment may result in a relatively favorable prognosis for neurological symptoms. Immunotherapy, such as plasma exchange, IVIg, and steroids, is also used for neurological symptoms,

but patients with antibody positive against the intracellular antigens, such as anti-Hu antibodies, are generally resistant to immunotherapy¹¹. It has also been reported that IVIg is less effective for central nervous system symptoms than for peripheral nervous system symptoms. In the present case, improvement in the tendon reflexes of the upper extremities was observed after IVIg administration, suggesting that IVIg improved the peripheral neuropathy¹². Syndromes mediated by surface neuronal autoantibodies usually respond to prompt immunotherapy in addition to oncological therapies; some syndromes associated with 'onconeural' antibodies may not respond, but there are important exceptions¹³.

In a study of 686 cases with metastatic cancer of unknown primary site¹⁴, bones, lungs, and liver were the common sites of metastases, whereas adenocarcinoma¹⁵, carcinoma, and anaplastic carcinoma were the most common histological types. Small cell carcinoma was relatively rare and found in only 3 cases (0.4%)¹⁴. In our patient with isolated para-aortic lymph node carcinoma, the repeated CT scan did not show a new primary lesion. The neurological symptoms persisted, but did not progress and the patient survived. If the metastatic lesion is single station lymph node carcinoma, treatment with excision alone may be sufficient and some patients survive for several years^{16,17}.

In conclusion, we reported a case of anti-Hu antibody positive PNS as one of the differential diagnoses of subacute progressive ABP. In such patients, various systemic diseases should be promptly excluded, and the tumor search should be repeated.

Table 2 Antiganglioside antibody results in the present case.

	IgM	PA +	IgG	PA +
GM1	-	-	-	-
GM2	-	-	-	-
GD1a	-	-	-	-
GD1b	-	-	-	-
GT1a	-	-	-	-
GT1b	-	-	-	-
GQ1b	-	-	-	-
GD3	-	-	-	-
GalNAc-GD1a	-	-	-	-
Gal-C	-	-	-	-
GA1	-	-	-	-

Results are displayed as the OD value based on the ELISA response. ELISA, enzyme-linked immunosorbent assay; OD, optic density; PA +, complex of glycolipid with phosphatidic acid.



Figure 1

Chest computed tomography showing enlargement of the isolated para-aortic lymph node (white arrow).

Tumor size; October 2020: 3.99 × 9.88 mm; November 2020: 6.79 × 14.76 mm; December 2020: 12.18 × 24.11 mm.

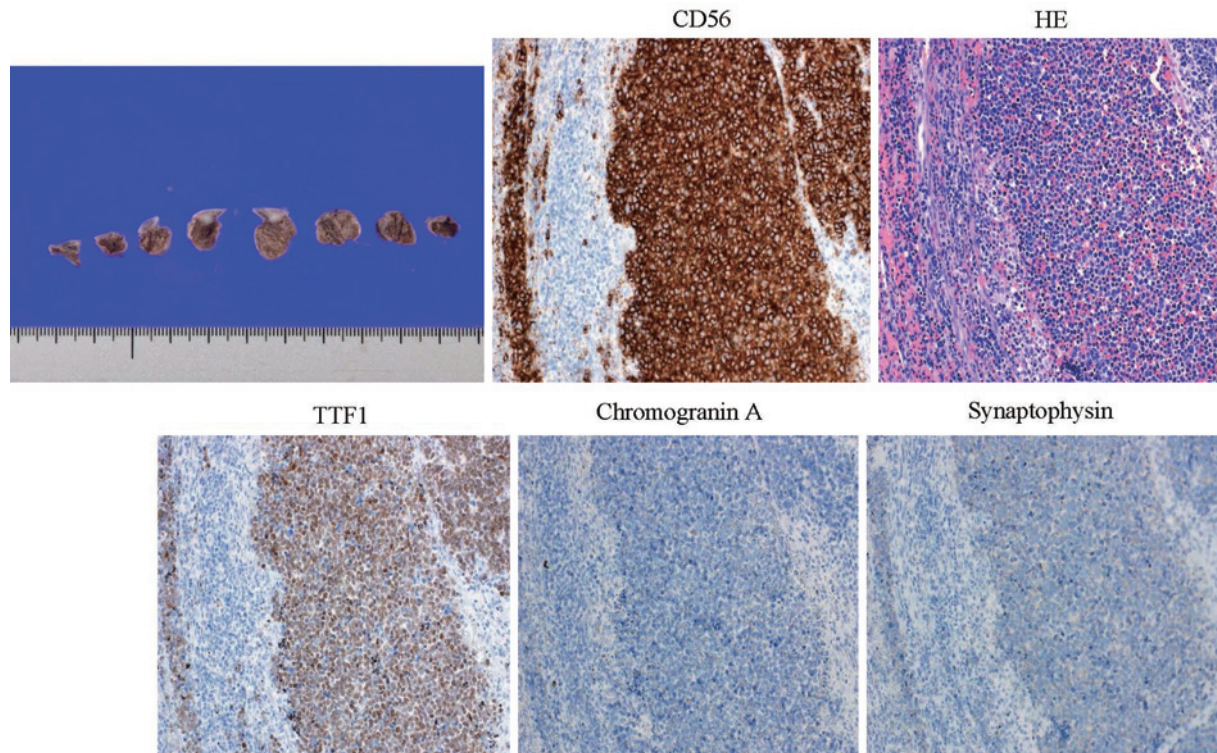


Figure 2

Tumor cells in the lymph nodes near the para-aortic arch were CD56-positive, TTF1-positive, chromogranin A-negative, and synaptophysin-weakly positive, which were consistent with small cell carcinoma.

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